

INHIBITION OF PHAGOCYTOTIC METABOLIC CHANGES OF LEUKOCYTES BY AN INTRACELLULAR  
CALCIUM-ANTAGONIST 8-(N,N-DIETHYLAMINO)-OCTYL-3,4,5-TRIMETHOXYBENZOATE

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Summary

The role of calcium in regulating the activity of leukocytes to generate and release superoxide was studied by using an intracellular calcium-antagonist, 8-(N,N-diethylamino)-octyl-3,4,5-trimethoxybenzoate. The antagonist inhibited the release of superoxide anions induced by a calcium-ionophore A23187 and the inhibition was relieved by the addition of calcium ions. The release induced by cytochalasin D or by the ingestion of bacteria was similarly inhibited by the calcium-antagonist. The result supports the hypothesis that an intracellular translocation of calcium is regulating the phagocytotic metabolic activity of leukocytes. The release of granule enzymes induced by the ionophore was also inhibited by the calcium antagonist.

Polymorphonuclear leukocytes produce superoxide anions ( $O_2^-$ ) and hydrogen peroxide during phagocytosis. These active oxygens are known to play an important role in bactericidal function of the cells (1-3). Several reagents, for example cytochalasin E (or D) (4,5) and a calcium ionophore A23187, also induce the metabolic change. The role of calcium in the activation of the oxygen metabolism has been discussed, based on the observations that the enhancement of the oxygen consumption was induced by the calcium-ionophore in the presence of  $Ca^{2+}$  (6) and the release of  $O_2^-$  induced by digitonin was inhibited by a calcium-chelator EGTA (7). If we assume the role of cytoplasmic  $Ca^{2+}$  as a second messenger to regulate the metabolism, it is necessary to consider not only the flux of  $Ca^{2+}$  through plasma membrane but also the possibility of an intracellular translocation of the ions between an intracellular storage pool and cytosol.

Abbreviation: TMB-8, 8-(N,N-diethylamino)-octyl-3,4,5-trimethoxybenzoate.

A new intracellular calcium antagonist 8-(N,N-diethylamino)-octyl-3,4,5-trimethoxybenzoate(TMB-8) has been synthesized. It blocks contraction of smooth and skeletal muscles induced by electrical stimulation, acetylcholine or caffeine, and the inhibition is relieved by extracellular  $\text{Ca}^{2+}$  (8,9). An application of this reagent in study of the ATP-secretion from blood platelets was reported: TMB-8 inhibited the secretion induced by either thrombin or the calcium-ionophore and the inhibition was overcome by extracellular  $\text{Ca}^{2+}$  (10).

In this paper, we report the inhibitory effect of TMB-8 on the activity of leukocytes to generate and release  $\text{O}_2^-$  and the reversal of its effect by the extracellular  $\text{Ca}^{2+}$ . The results suggest that an intracellular translocation of  $\text{Ca}^{2+}$  is regulating the phagocytotic metabolic changes of leukocytes.

#### METHODS

Polymorphonuclear leukocytes were those of guinea pig peritoneal exudates induced by a caseinate solution (5). The cells were suspended in a calcium-free Tris-Ringer solution (120 mM-NaCl, 5 mM-KCl, 1.2 mM-MgCl<sub>2</sub>, 20 mM-Tris-HCl buffer, pH 7.4).  $\text{CaCl}_2$  was added as described in Text. The activity to release  $\text{O}_2^-$  was assayed by the reduction of cytochrome c essentially as described previously (4). The reduction of cytochrome c was measured by a dual-wavelength spectrophotometer (Hitachi, 556) at 550-540 nm with a molar absorption coefficient of  $19.1 \times 10^3 \text{ M}^{-1} \text{ cm}^{-1}$ . The release of enzymes from granules was studied as described previously (5). The activities of myeloperoxidase, lysozyme,  $\beta$ -glucuronidase and lactate dehydrogenase were assayed by reported procedures (11-14). Superoxide dismutase was prepared from bovine erythrocytes by the method of McCord and Fridovich (15). Cytochalasin D, cytochrome c and phenolphthalein  $\beta$ -glucuronide were purchased from Sigma. Ionophore A23187 and TMB-8 were generous gifts of Eli Lilly Co., and Dr. Shoji Kume, Department of Medicine, Tokyo University, respectively. A23187 and TMB-8 were dissolved in dimethylsulfoxide.

#### RESULTS AND DISCUSSION

The addition of a calcium-ionophore A23187 to leukocyte suspension induced the release of superoxide anions even without the addition of  $\text{Ca}^{2+}$  to the medium, which was indicated by a continuous reduction of cytochrome c (Fig. 1, curve a). The reduction could be stopped by the addition of superoxide dismutase. The release was inhibited by an intracellular calcium-antagonist TMB-8, as shown by the curve b and was restored by the subsequent addition of  $\text{Ca}^{2+}$ , as shown by the curve c.

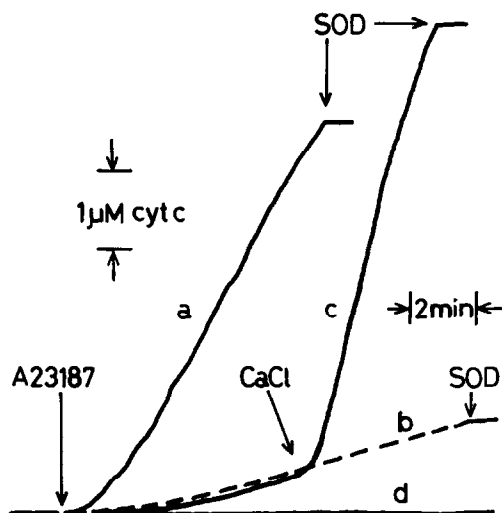


Fig. 1. The release of  $O_2^-$  from leukocytes treated with a ionophore A23187. The cells ( $2 \times 10^6$ ) in 1 ml of the  $Ca^{2+}$ -free Tris-Ringer solution containing 2 mM glucose and 50  $\mu$ M cytochrome c were incubated at 37°C with or without TMB-8 (100  $\mu$ M). The reaction was started by the addition of 5  $\mu$ M A23187 and the absorbancy change (550-540 nm) was recorded. The reduction of cytochrome c was completely inhibited by 10  $\mu$ g of superoxide dismutase (SOD). Curves: a) without TMB-8, b) with TMB-8, c) with TMB-8 and subsequently with  $CaCl_2$ , d) control without A23187.

The effect of extracellular  $Ca^{2+}$  concentration on the release of  $O_2^-$  induced by A23187 was different in the presence and absence of TMB-8 (Fig. 2a). The activity of the cells to release  $O_2^-$  was essentially proportional to extracellular  $Ca^{2+}$  concentrations when the cells were treated with TMB-8, whereas the activity of the cells not treated with TMB-8 was apparently independent of extracellular  $Ca^{2+}$  when the  $Ca^{2+}$  concentration was low but it increased linearly with the  $Ca^{2+}$  concentration at higher  $Ca^{2+}$  concentrations. The release in the absence of TMB-8 and at low concentration of extracellular  $Ca^{2+}$  may be ascribed to the effect of A23187 to accelerate the release of  $Ca^{2+}$  from an intracellular storage granule to the cytosol, as suggested by Charo et al. (10), whereas the release of  $O_2^-$  at higher concentration of extracellular  $Ca^{2+}$  may essentially be due to the transport function of the ionophore through plasma membrane. Thus, the percentage of the inhibition caused by TMB-8 at a given concentration of extracellular  $Ca^{2+}$  was dependent on the  $Ca^{2+}$  concentration

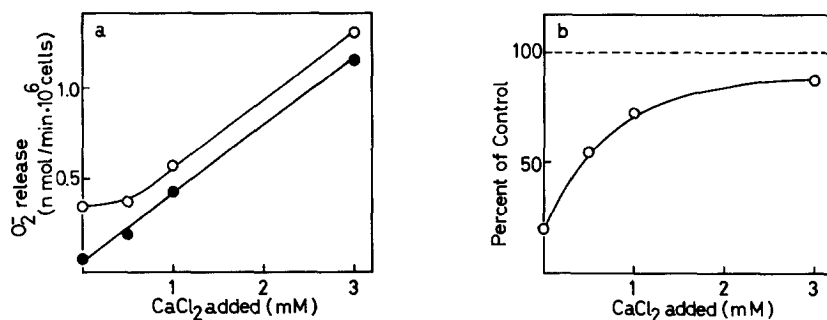


Fig. 2a. The effect of added  $Ca^{2+}$  concentration on the  $O_2^-$  release induced by A23187 in the presence and absence of TMB-8. The conditions were the same as described for Fig. 1. (●) with TMB-8, (○) without TMB-8.

Fig. 2b. The activity of the cells treated with TMB-8 as percent of the control activity without TMB-8 at a given concentration of  $Ca^{2+}$  (from Fig. 2a).

(Fig. 2b): the inhibition was strong in  $Ca^{2+}$ -free medium and it decreased gradually with the increase of extracellular  $Ca^{2+}$  concentration. This observation may support another hypothesis of Charo et al. (10) that TMB-8 serves as a general antagonist of intracellular  $Ca^{2+}$  by inhibiting the release of  $Ca^{2+}$  from a storage site.

The inhibition of the  $O_2^-$ -release by TMB-8 was not limited to the activity induced by the calcium-ionophore. As shown in Table I, the  $O_2^-$ -release induced by the ingestion of heat-killed *E. coli* or by the interaction of cytochalasin D was similarly inhibited by TMB-8. This suggests the general role of intracellular  $Ca^{2+}$  in the regulation of the  $O_2^-$ -release reaction. Verapamil which

Table I. The effect of TMB-8 on the  $O_2^-$ -release activity induced by *E. coli*, cytochalasin D or A23187.

	Activity (nmol/min per $10^6$ cells)		
	<i>E. coli</i> (0.9 mg)	cytochalasin D (10 $\mu$ g)	A23187 (5 nmol)
Control	0.84	1.20	2.49
TMB-8 50 $\mu$ M	0.49	0.40	2.49
100 $\mu$ M	0.42	0.13	1.06
200 $\mu$ M	0.14	0	0.51
300 $\mu$ M	0.04	0	0.07

The conditions were essentially the same as described for Fig. 1. The inducers were added 5 min after the preincubation with or without TMB-8.

Table II. The effect of TMB-8 on the release of granule enzymes induced by A23187

	Activities of enzymes released (% of total activity)					
	Myeloperoxidase		Lysozyme		$\beta$ -glucuronidase	
	Control	TMB-8	Control	TMB-8	Control	TMB-8
Control	3.5		4.1		3.4	
A23187						
$\text{Ca}^{2+}$ -free	16.2	9.3	14.5	9.3	8.3	7.8
3 mM- $\text{Ca}^{2+}$	59.8	52.1	29.2	21.2	17.3	16.6

Leukocytes ( $4 \times 10^7$ ) in 2 ml of the Tris-Ringer solution were preincubated for 5 min at  $37^\circ\text{C}$  with or without TMB-8 (200  $\mu\text{M}$ ) and the reaction was started by A23187 (10  $\mu\text{M}$ ). After 10 min, the samples were cooled in ice and the supernatant was separated by centrifugation at  $900 \times g$  for 10 min at  $4^\circ\text{C}$ . The activity in the supernatant was expressed as the percent of total activity obtained by the disruption of the cells with 0.2 % Triton X-100. No changes in the release of lactate dehydrogenase was observed by the treatments.

has been used to inhibit the influx of extracellular  $\text{Ca}^{2+}$  (16) showed only 20 % inhibition at 10  $\mu\text{M}$  (induction by cytochalasin D at 1 mM- $\text{Ca}^{2+}$ ).

TMB-8 also inhibited the release of granule enzymes from the cells (Table II). Myeloperoxidase and lysozyme which are known to be localized in granules, were released from the cells into incubation medium, when the cells were treated with the ionophore A23187, while the release of lactate dehydrogenase, a cytosol enzyme, was essentially unaffected by the treatment. The release of the granule enzymes was accelerated by the addition of  $\text{Ca}^{2+}$  ions and was inhibited by TMB-8. The inhibition by TMB-8 was strong in calcium-free medium, whereas it was not so apparent in the presence of the added  $\text{Ca}^{2+}$ . The situation was very much similar to that of the  $\text{O}_2^-$ -release and a similar mechanism is supposed to be operating in the regulation of the both processes. The effect of TMB-8 on the release of another granule enzyme,  $\beta$ -glucuronidase, was not so clear. This may be a reflection of the different localization of myeloperoxidase and  $\beta$ -glucuronidase in guinea pig leukocytes (11). Lysozyme may distribute in the both kinds of granules as in the case of human leukocytes (17) and it may explain the intermediary properties of lysozyme between myeloperoxidase and  $\beta$ -glucuronidase.

There is a general agreement that calcium is a second messenger of several stimulus-secretion or stimulus-contraction coupling processes. Intracellular free calcium can be regulated either by an influx of extracellular  $\text{Ca}^{2+}$  or by an intracellular translocation of the ions, or by both, but an importance of the intracellular process in short-time metabolic changes has been suggested for various kind of cells (18). The present observation, together with accumulated evidences in the researches of other cells, may allow us to speculate that an intracellular translocation of  $\text{Ca}^{2+}$  is playing an important role in the regulation of phagocytotic metabolism in polymorphonuclear leukocytes.

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#### REFERENCES

1. Iyer, G.Y.N., Islam, M.F., and Quastel, J.H. (1961) *Nature*, **192**, 535-541
2. Holmes, B., Page, A.R., and Good, R.A. (1967) *J.Clin.Invest.* **46**, 1422-1432
3. Babior, B.M., Kipnes, R.C., and Curette, J.T. (1973) *J.Clin.Invest.* **52**, 741-744
4. Nakagawara, A., and Minakami, S., (1975) *Biochem.Biophys.Res.Comm.* **64**, 760-767
5. Nakagawara, A., Shibata, K., Takeshige, K., and Minakami, S., (1976) *Exp.Cell Res.* **101**, 225-234
6. Romeo, D., Zabucchi, G., Miani, N., and Rossi, F., (1975) *Nature*, **253**, 542-544
7. Cohen, H.J., and Chovaniec, M.E. (1978) *J.Clin.Invest.* **61**, 1088-1096
8. Malagodi, M.H., and Chiou, C.Y. (1974) *Eur.J.Pharmacol.* **27**, 25-33
9. Malagodi, M.H., and Chiou, C.Y. (1974) *Pharmacology* **12**, 20-31
10. Charo, I.F., Feinman, R.D., and Detwiler, T.C. (1976) *Biochem.Biophys.Res.Comm.* **72**, 1462-1467
11. Mitchell, R.H., Karnovsky, M.J., and Karnovsky, M.L. (1970) *Biochem.J.* **116**, 207-216
12. Smolelis, A.N., and Hartsell, S.E. (1949) *J.Bacteriol.*, **58**, 731-736
13. Hirschhorn, R., Hirschhorn, K., and Weissmann, G. (1967) *Blood*, **30**, 84-102
14. Bergmeyer, H.U., and Bernt, E. (1974) *Method in Enzymatic Analysis*, Academic Press, New York, pp. 574-579
15. McCord, J.M., and Fridovich, I. (1969) *J.Biol.Chem.* **244**, 6049-6055
16. Davis, G., Somers, G., and Malaisse, W.J. (1975) *Biochem.Biophys.Res.Comm.* **67**, 525-529
17. Bretz, U., and Baggiolini, M. (1974) *J.Cell Biol.* **63**, 251-269
18. Rasmussen, H., and Biale, D.D. (1975) *Calcium Transport in Contraction and Secretion*, North-Holland Pub.Comp., Amsterdam, pp. 111-121